

8-METHOXYNEORAUTENOL AND RADICAL SCAVENGING FLAVONOIDS FROM *ERYTHRINA ABYSSINICA*

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ABSTRACT. A new pterocarpan (named 8-methoxyneorautenol) was isolated from the acetone extract of the root bark of *Erythrina abyssinica*. In addition, the known isoflavonoid derivatives eryvarin L, erycristagallin and shinpterocarpin were identified for the first time from the roots of this plant. The structures were determined on the basis of spectroscopic evidence. The new compound showed selective antimicrobial activity against *Trichophyton mentagrophytes*. The acetone extract of the root bark of *E. abyssinica* showed radical scavenging activity towards 2,2-diphenyl-1-picrylhydrazyl radical (DPPH). The pterocarpenes, 3-hydroxy-9-methoxy-10-(3,3-dimethylallyl)pterocarpene and erycristagallin, were the most active constituents of the roots of this plant and showing dose-dependent activities similar to that of the standard quercetin.

KEY WORDS: *Erythrina abyssinica*, Root bark, Leguminosae, Pterocarpan, 8-Methoxyneorautenol, Pterocarpenes, 3-Hydroxy-9-methoxy-10-(3,3-dimethylallyl)pterocarpene, Erycristagallin, Antimicrobial, Radical scavengers, DPPH

INTRODUCTION

Erythrina abyssinica (Leguminosae) is distributed in warm regions of southern Africa and the savannah of eastern Africa [1]. It is used in traditional medicine for treatment of malaria, elephantiasis, trachoma and syphilis [2]. Previous phytochemical studies on *E. abyssinica* have shown that the plant elaborates alkaloids [1], flavanones, pterocarpanes, chalcones and isoflavonoids [1, 3]; some of which have been shown to have antimicrobial [2, 4], anti-plasmodial [5, 6] and inhibitory effects on protein tyrosine phosphatase 1B (PTP1B) [3] activities. In this paper, we report on the isolation and structure elucidation of a new pterocarpan, along with known isoflavonoid derivatives. The radical scavenging activities of the acetone extracts of the roots and stem bark of *Erythrina abyssinica*, and some of the compounds isolated from these extracts are also reported.

RESULTS AND DISCUSSION

The HRMS analysis of compound **1** showed $[M+1]^+$ peak at m/z 353.1397 corresponding to the molecular formula of $C_{21}H_{20}O_5$. The 1H and ^{13}C NMR spectra (Table 1) showed this compound to be a pterocarpan derivative. Furthermore the presence of hydroxyl, methoxyl and a 2,2-dimethylpyran functionalities was established from NMR and MS data. In the A ring, the presence of two singlet aromatic protons at δ 7.12 (for H-1) and δ 6.37 (for H-4) requires that C-2 and C-3 are substituted. The HMBC correlation of one of the olefinic protons (H-4') of the 2,2-dimethylpyran functionalities with C-1 is consistent with this group being at C-2/C-3 with the expected oxygenation at C-3.

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Table 1. ^1H (500 MHz) and ^{13}C NMR (125 MHz) data along with HMBC correlations in **1** (CDCl_3).

Position	1		HMBC (2J , 3J)
	δ_{C}	δ_{H} (J in Hz)	
1	128.3	7.12 s	C-3, 4a, 11a, 4'
2	116.4		
3	154.6		
4	104.7	6.37 s	C-2, 3, 4a
4a	156.2		
6	66.5	3.62 dd (-10.9, 11.3) 4.22 dd (5.1, -10.9)	C-6a, C-4a, 6a, 11a
6a	40.3	3.49 ddd (5.1, 6.4, 11.3)	C-6, 11a
6b	116.9		
7	107.8	6.79 s	C-6a, 8, 9, 10a
8	141.1		
9	146.6		
10	98.1	6.51 s	C-8, 9, 10a, 6b
10a	153.9		
11a	78.2	5.42 d (6.4)	C-1, 4a, 6
11b	112.4		
2'	76.6		
3'	129.1	5.54 d (9.8)	C-2, 2'
4'	121.6	6.31 d (9.8)	C-1, 3, 2'
2' (CH_3) ₂	27.9	1.40 s	
	28.2	1.43 s	
8-OCH ₃	57.0	3.86	C-8

In the D-ring the presence of two *para*-oriented aromatic protons at δ 6.79 (H-7) and 6.51 (H-10) and the chemical shift values of the aromatic carbon atoms, δ 116.9 (C-6b), 107.8 (C-7), 141.1 (C-8), 146.6 (C-9), 98.1 (C-10) and 153.9 (C-10a) require that the methoxyl and the hydroxyl groups are substituted at C-8 and C-9, respectively. The position of the methoxyl group was fixed at C-8 rather than C-9, from NOEDIFF experiment; where irradiation of the methoxyl group showed NOE interaction on H-7 (δ 6.79). This was confirmed through the HMBC correlation between the methoxyl protons and C-8, and also between H-7 and C-8 (Table 1). This compound was therefore assigned structure **1** for which the trivial name 8-methoxyneorautenol was suggested. The high negative optical rotation ($[\alpha]_{\text{D}} -246.6^\circ$) is consistent with 6*aR*:11*aR* configuration in this compound [7].

In addition, the pterocarpan shinpterocarpin [8, 9] and the arylbenzofuran eryvarin L (**2**) have been identified from the roots of this plant for the first time. Compound **2** is rare and has only been reported from the twigs of *E. abyssinica* [1] and the roots of *E. varigata* [10]. It is worth noting that this compound co-occurs in *E. varigata* with the isoflav-3-ene eryvarin H (**3**) which has identical oxygenation pattern as compound **2** [10]. It has been proposed that eryvarin L is derived from eryvarin H in this plant [11]. These compounds also co-occur in *E. abyssinica* [5], supporting the biogenetic relationship between arylbenzofurans and isoflav-3-enes. The arylbenzofuran burttinol D co-occurs with the isoflav-3-ene burttinol A in the roots of *E. burttii* [12], and similar biogenetic relationship has been suggested for these compounds [11].

Compound **1** showed selective but weak antimicrobial activity against *Trichophyton mentagrophytes* with MIC value of 45 $\mu\text{g}/\text{mL}$ [4], while it was inactive against a range of microorganisms [*vis a vis* *Staphylococcus aureus* (ATCC 25923), Methicillin resistant *Staphylococcus aureus* (clinical isolate), *Escherichia coli* (ATCC 25922), *Pseudomonas aeruginosa* (ATCC 27853), *Candida albicans* (ATCC 90028), *Trichophyton mentagrophytes*, *Cryptococcus neoformans* (ATCC 66031) and *Microsporum gypseum* (clinical isolate)].

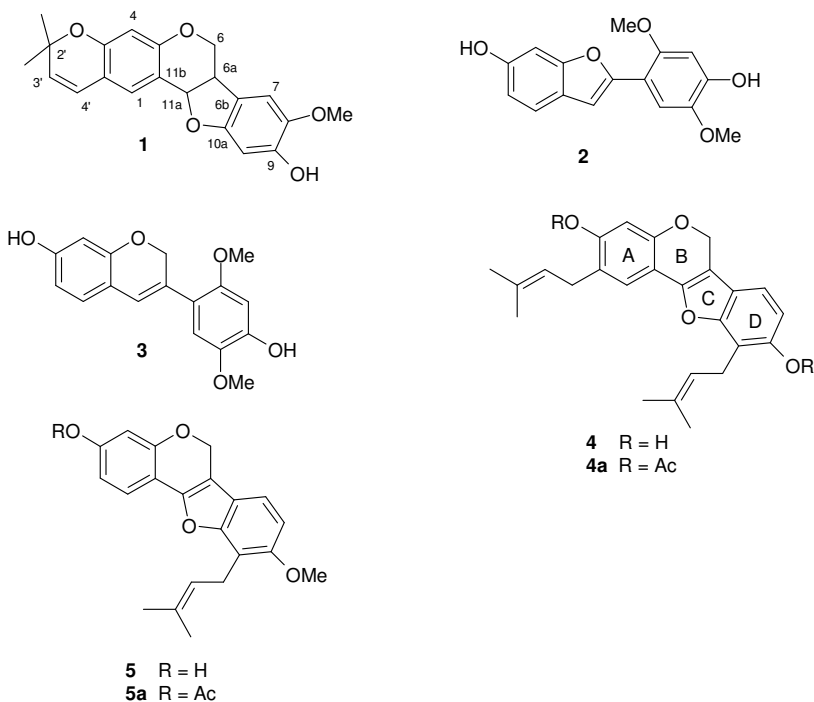
Preliminary tests for radical scavenging activities (RSA), using 2,2-diphenyl-1-picrylhydrazyl (DPPH) free radical as a spray reagent on TLC plates, of the crude acetone

extract of the root bark and the stem bark of *E. abyssinica* indicated the presence of active compounds. Using the spectrophotometric method, the RSA of the crude extracts were determined and both extracts showed potent activities ($EC_{50} = 7.7$ and $17.4 \mu\text{g/mL}$ for root bark and stem bark extracts, respectively).

Some of the compounds previously isolated from the root bark [5] and the stem bark [6] of *E. abyssinica* were tested. Activities were observed among flavanones, pterocarpenes and isoflav-3-enes (Table 2). Among these isolates, the pterocarpenes erycristagallin (**4**) and 3-hydroxy-9-methoxy-10-(3,3-dimethylallyl)pterocarpene (**5**) were identified as the most active principles in this plant. The activities of these pterocarpenes were tested at different concentrations (Figure 1) and both showed potent (EC_{50} $8.2 \mu\text{M}$ for **4** and $10.8 \mu\text{M}$ for **5**) and dose dependent radical scavenging activities towards DPPH, similar to that of the standard quercetin (EC_{50} value $5.4 \mu\text{M}$).

Table 2. Radical scavenging activities towards DPPH of compounds from *E. abyssinica*.

Compound	EC_{50} (μM)
Abyssinone IV	32.4
Abyssinone V	30.1
Abyssinin III	21.7
4	8.2
5	10.8
4',7-Dihydroxy-2',5'-dimethoxyisoflav-3-ene	62.0
Quercetin	5.4



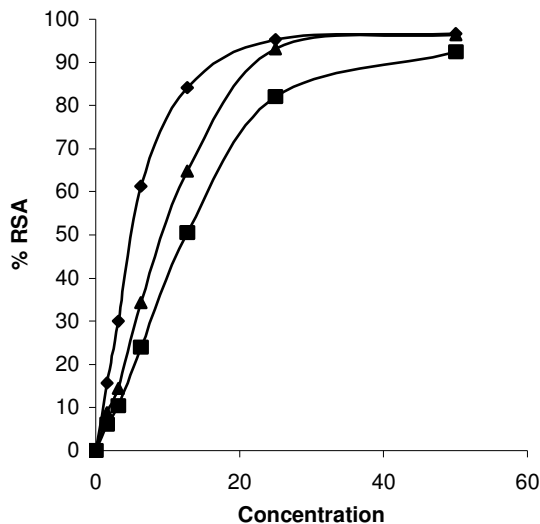


Figure 1. Radical scavenging activities (RSA) of different concentrations (in μM) of **4** (■) and **5** (▲) together with quercetin (◆) against DPPH.

As the standard quercetin, these pterocarpenes have free phenolic group(s) required for activity; and as expected [13], the acetate derivatives **4a** and **5a** were inactive even at $50 \mu\text{M}$. The structural requirements for the high radical scavenging activity of quercetin and other related flavonoids has been reported to be due to the O-dihydroxyl structure in B-ring; the 2,3 double bond in conjunction with the 4-oxo function in the C-ring; and the 3- and 5-hydroxyl groups with hydrogen bonding to the keto group [14]. The structural features in the pterocarpenes, however, do not satisfy these requirements and yet these compounds showed very high radical scavenging activities. In both pterocarpenes where all four rings (A-, B-, C- and D-rings) are coplanar, and the double bond in C-ring appears to play an important role for the high radical scavenging activity, extending the conjugation among the four rings to stabilize the radical formed after loss of hydrogen to DPPH. The presence of 3,3-dimethylallyl group(s) in these pterocarpenes not only enhances the radical scavenging activities, but also increase the lipophilicity of these compounds and hence probably make them protect low density lipoproteins (LDL) better than the more polar flavonoids such as quercetin [13].

EXPERIMENTAL

General

Analytical TLC: Merck pre-coated silica gel 60F₂₅₄ plates. CC on oxalic acid impregnated silica gel 60 (70-230 mesh). EIMS: direct inlet, 70 eV, on a SSQ 710, Finnigan MAT mass spectrometer. HRMS were recorded with electrospray ionisation on a ESI-QTOFmicro mass spectrometer from Waters Inc. ¹H NMR (500 or 200 MHz) and ¹³C NMR (125 or 50 MHz) were recorded on Bruker or Varian-Mercury spectrometers using TMS as internal standard. HMQC and HMBC spectra were acquired using the standard Bruker software.

Plant material

The stem bark and root bark of *E. abyssinica* were collected near Thika town, Kenya, in December 2006. The plant was identified at the University Herbarium, Botany Department, University of Nairobi, where a voucher specimen is deposited.

Extraction and isolation

Air-dried and ground root bark (1.7 kg) of *E. abyssinica* was extracted with acetone by cold percolation at 25 °C. Removal of the solvent under reduced pressure afforded a brown sticky extract (109 g). A 100 g portion of the extract was fractionated on oxalic acid-impregnated silica gel (350 g) eluting with n-hexane containing increasing amounts of dichloromethane (5, 10, 20, 30, 40, 50, 60, 70, 80, 90 and 100 %). The fractions eluted with 20 % and 30 % CH₂Cl₂ were combined and further subjected to CC on oxalic acid-impregnated silica gel (eluting with 2 % EtOAc in n-hexane) and purified on Sephadex LH-20 (eluted with CH₂Cl₂/MeOH; 1:1) to give compound **1** (9 mg). The fraction eluted with 50 % CH₂Cl₂ in hexane was purified on Sephadex LH-20 (eluted with CH₂Cl₂/MeOH; 1:1) and then by preparative TLC (silica gel, hexane/CH₂Cl₂, 3:7) to give shinpterocarpin (6 mg) and **4** (43 mg) and **5** (26 mg). The fraction eluting with 70 % CH₂Cl₂ in n-hexane was purified on a preparative TLC (n-hexane/CH₂Cl₂, 1:4) to give compound **2** (7 mg).

8-Methoxyneorautenol (1)

Amorphous powder. $[\alpha]_D^{25}$ -246.6° (c 0.01, MeOH). UV λ_{max} (MeOH): 283, 234 nm. IR ν_{max} : 3580, 2825, 1620, 1610, 1596, 1495 cm⁻¹. ¹H NMR (Table 1). ¹³C NMR (Table 1). ESI-HRMS m/z [M+1]⁺ 353.1397 C₂₁H₂₁O₅. EIMS m/z (rel. int.): 352 (46, [M]⁺), 337 [M-Me]⁺ (100).

Radical scavenging activity test using DPPH

To a methanolic solution (3 mL) of DPPH (100 μM), 0.5 mL of each of the test compound from a 50 μM solution (10 μg/mL for crude extract) were added and the mixture was shaken and left to stand for 30 min. The radical scavenging activities were estimated as the percent decrease of the absorbance of DPPH (100 μM) at 517 nm [15]. Tests were done at six different concentrations (50, 25, 12.5, 6.25, 3.13 and 1.56 μM). In all cases the mean values were used from triplicate experiments. EC₅₀ values were calculated using Finney's probit analysis for quantal data [16].

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